

OXIDATIVE PREPARATION OF OPTICALLY ACTIVE N-HYDROXY- α -AMINO ACID AMIDES

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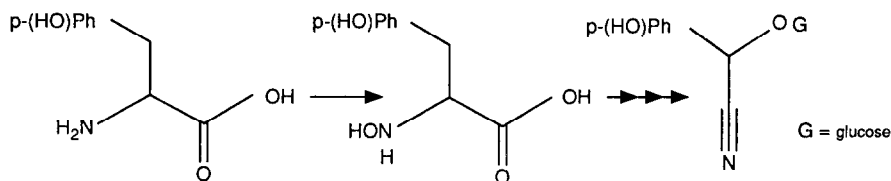
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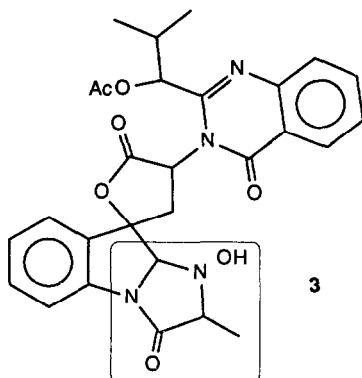
Summary Two routes are presented for the conversion of optically active α -amino acid amides into the title compounds. One route (route A) features the formation of the Schiff's base **6** which is subsequently oxidized to the corresponding oxazindines **7**. Route B is characterized by the formation of an imidazolium **11** which is hydroxylated to compound **12**. Alcoholysis of **7** and **12** in the presence of hydroxylamine hydrochloride yields the title compounds in overall yields ranging from 65 to 85% (route A) and from 14 to 21% (route B).

Introduction

N-hydroxy- α -amino acid derivatives are widely encountered in nature¹. They can be found amongst others as constituents of peptides, to which physiological properties can be attributed like antibiotic activity² for example. Moreover, it has been postulated³ that N-hydroxy amino acids play an important role in the metabolism of peptidogenic amino acids. For example in the biosynthesis of dhurrin⁴, the intermediate N-hydroxy tyrosin **1** has been proposed (scheme 1). Another natural product that contains an N-hydroxy amino acid as a structural feature is nortryptoquinoline **3**, a toxic metabolite^{5a} isolated from a strain of *Aspergillus clavatus*.



scheme 1



Most of the methods reported for the synthesis of N-hydroxy- α -aminoacids yield racemic mixtures^{5b} Hence, there is a need for a general route to homochiral N-hydroxy- α -amino acids

Our contribution to the answer addressing this challenge has resulted in three approaches The first one features a substitution reaction involving triflates of α -hydroxy esters and hydroxylamine or derivatives thereof⁶ The second approach is based on the enzymatic resolution of N-benzyloxy-amino acid ethyl esters⁷ Here we report our third approach which is based on the selective N-oxidation of derivatives of optically active amino acids

Although oxidation of amino acids directly to the title compounds seems to be straight forward, a method for the *direct* oxidation of the amino function in amino acids has been unsuccessful⁸

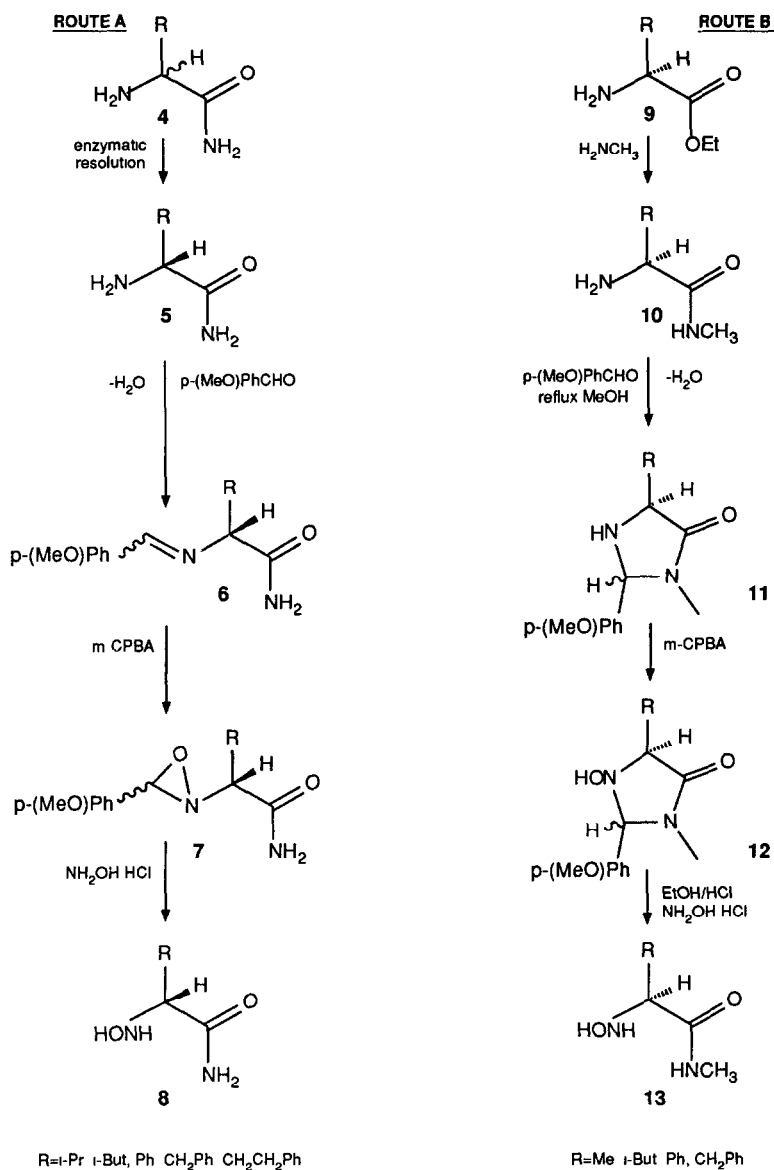
Indirect oxidation - of which two examples are discussed here - however offers a viable approach Połoński *et al*⁹ demonstrated that conversion of the amino function of an α -amino acid ester into an imine renders this functionality susceptible to N-hydroxylation, a process involving an oxaziridine as intermediate (*cf* structure 7, scheme 3) Despite later improvements to this method¹⁰, this approach still suffers from variable yields that are - occasionally - unacceptably low We now report that this approach can be made more efficient and reliable by employing α -amino carboxy amides 5 (scheme 2, route A)

Furthermore, from Buchi's synthesis^{5a} of nortryptoquivaline 3 we concluded that N-hydroxylation should also be feasible when a secondary amine is subjected to oxidation conditions Studies on the possibility whether the imidazolidinones 11 - masked derivatives of the corresponding L-amino acid amides 10 - could be oxidized to the 1-hydroxy-imidazolidinones 12 as potential precursors for 13, have proved route B (scheme 2) to be viable indeed

Results and discussion

Route A

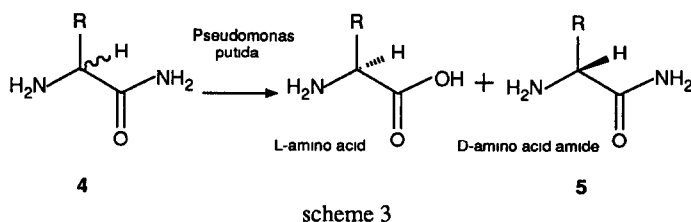
Optically active amino acid amides 5 are readily available on a large scale by applying enzymatic hydrolysis to D,L-amino acid amides¹¹ 4 Using an L-specific aminopeptidase from *Pseudomonas putida*, stereoselective hydrolysis of the L-amino acid amide into the L- α -amino acid is achieved while the D-amino acid amide 5 remains untouched, see scheme 3 Separation of the



scheme 2

ester and the amide is afforded by adding one equivalent (with respect to the D- α -amino acid amide) of an aromatic aldehyde, e.g. anisaldehyde, to the enzymatic hydrolysate¹². Since the Schiff's base **6** of the amino acid amide, which is formed quantitatively, is insoluble in water it can easily be isolated by filtration.

The Schiff's base of the amino acid derivative having either the L or D chirality is the intermediate of choice for the oxidation procedure.



The dry imine **6** when dissolved in dry dichloromethane at -15°C is nearly quantitatively converted into the oxaziridine **7** when a slight excess of *m*-CPBA is used. Subsequent treatment with hydroxylamine gives the hydrochloride of the N-hydroxy α -D- or L-amino acid amide **8**. Trituration with ether gives white crystals in good yields based on L- or D- α -amino acid amide **5**, see table 1.

Table 1 Chemical yields of the conversion **5** \rightarrow **8** (route A)

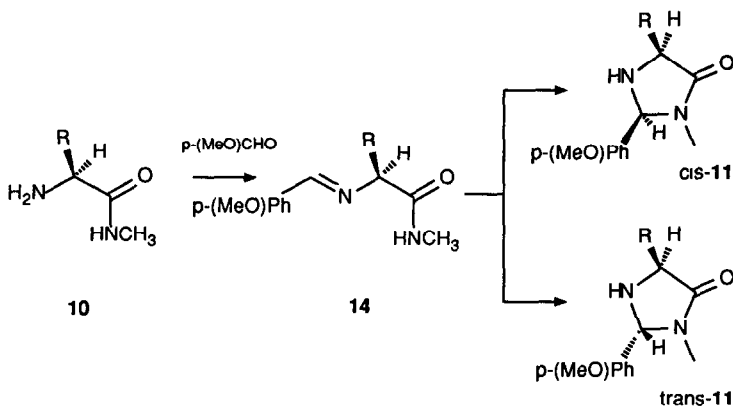
entry	R	chemical yield (%)	
		5 \rightarrow 6	6 \rightarrow 8
a	<i>i</i> -Pr	≥ 95	76
b	<i>i</i> -But	≥ 95	65
c	Ph	≥ 95	83
d	CH_2Ph	≥ 95	80
e	$\text{CH}_2\text{CH}_2\text{Ph}$	≥ 95	80

To establish the stereochemical identity, the N-hydroxy- α -amino acid amides **8** are reduced in a Parr-apparatus with Pd/C. The specific rotations of the resulting α -amino acid amides **5** were in good agreement with those of the starting material **5**.

Route B

Subsequently, we studied the oxidation of imidazolidinones **11**. The latter compounds are easily obtained by refluxing a solution of the amide **10** and anisaldehyde in methanol. For this cyclization reaction an aldehyde was selected and not a ketone as we observed that the animals resulting from ketones, *e.g.* acetone, were very difficult to hydrolyse after the oxidation step. From the reaction with *p*-methoxybenzaldehyde two diastereomers emerge, the ratio of which in some cases could be determined by isolation of the separate diastereomers, see scheme 4.

The total yields of the two diastereomers of **11** together average 75% after purification by column chromatography, see table 2. It should be stressed here that reaction of **10** with the aldehyde yields the five membered ring only at elevated temperature, at room temperature the corresponding Schiff's base **14** is formed as discussed for route A. Refluxing in MeOH probably causes the initially formed Schiff's base **14** to cyclize to give **11**.



scheme 4

The principle of the conversion $14 \rightarrow 11$ has a precedent in literature¹³

The oxidation of **11** to **12** is performed as before with one equivalent of *m*-chloroperbenzoic acid in methanol. Subsequently, the ring is cleaved by treatment with ethanolic HCl and an equimolar amount of H₂NOH HCl (see scheme 2).

The solvolysis of **12** by ethanolic HCl alone also takes place, but recondensation of the N-hydroxy amino acid amide **13** with the released aldehyde moiety to give the corresponding nitron decreases the yield of **13**. This problem is solved by the addition of H₂NOH HCl which binds the aldehyde diethyl acetal liberated from the solvolysis reaction.

Table 2 Chemical yields of the conversion $9 \rightarrow 13$ (route B)

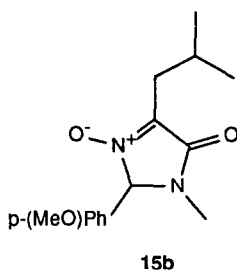
entry	R	chemical yield (%)			
		9→10	10→11 ^a	11→12 ^a	12→13
a	Me	99	36/36 ^b	24 ^c	78
b	<i>i</i> -But	93	38/38 ^d	9/27 ^e	78
c	Ph	99	50/26 [#]	--/88	37 ^f
d	CH ₂ Ph	99	46/33 [#]	--/74	83

^a) diast I/diast II ^b) ratio estimated from ¹H-NMR ^c) one diastereomer isolated ^d) ratio estimated from TLC ^e) a mixture of diastereomers of **11b** was oxidized, but the separate diastereomers of **12b** could be isolated ^f) yield not optimized [#]) diastereomers isolated in the ratio given

From table 2 it can be seen that the yield of the oxidation step $11 \rightarrow 12$ varies with the nature of the side chain and is highest when R = phenyl or benzyl (entries c and d). Another feature is that one of the diastereomers of **11a-d** is oxidized significantly faster than the other. In the case of **11b** the reaction with the slow reacting diastereomer is accompanied by the formation of more side

products When the slower reacting diastereomers of **11c** and **11d** were treated with more than one molar equivalent of *m*-CPBA, the corresponding N-hydroxylated compound **12** could not be isolated

The formation of **12b** was also accompanied by a small amount of the corresponding, overoxidized product **15b**



The optical purity(100%) of **13a** could be determined by comparison to a reference compound

Conclusions

Two routes for the synthesis of optically pure N-hydroxy- α -amino acid amides are described and in both routes the stereochemical identity of the starting material is retained

Route A starts with the optically pure amino acid amides **5** which are available in large quantities by methods developed at DSM¹¹ This route has proven to be very efficient (overall chemical yields ranging from 65 to 85%) and yields the chiral N-hydroxy- α -amino acid amides **8** Although the yield of the oxidation step **6** \rightarrow **7** has not been determined separately it can be concluded that this reaction proceeds in high yield(>65%)

Route B starts with the amino acid amides **10** and follows the reaction sequence **10** \rightarrow **11** \rightarrow **12** \rightarrow **13** which shows some noteworthy features In the conversion **10** \rightarrow **11** diastereoselectivity is nearly absent In the conversion **11** \rightarrow **12** only one diastereomer is oxidized cleanly, which drastically reduces the total yield of this oxidation step In route B the presence of an aromatic side chain increases the yield of the oxidation reaction In route A this reaction invariably proceeds in good yield regardless of the substituent present The alcoholysis of **7** and **12** carried out in the presence of H₂NOH HCl yields the desired title compounds in satisfactory yields

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Experimental part

¹H-NMR spectra were measured on a Bruker WH-90 spectrometer Infra red spectra were measured on a Perkin Elmer 298 spectrometer Mass spectra were obtained with a double focussing VG 7070E spectrometer Optical rotations were taken on a Perkin Elmer 241 polarimeter Thin-layer chromatography(TLC) was carried out by using Merck precoated silicagel F-254 plates(thickness 0.25 mm) For preparative column chromatography Merck silicagel type H60 was used

General procedure for the preparation of the Schiff's bases 6

Solution of 5-10 % w/w of the amides **5** are made in water of 40 °C (5 % w/w for R= Ph, and 10 %w/w for R = *i*-Pr)

The pH of the solution is adjusted to 11 by adding 1 N KOH

Then in about 15 min 1-10 equivalents of *p*-methoxybenzaldehyde is added to the solution After 2 hours of stirring at roomtemperature the cristallized Schiff's bases are isolated by filtration, washed with water and dried *in vacuo* The products **6** are obtained in nearly quantitative yields

General procedure for the preparation of oxaziridines 7

100 mmol of the Schiff's bases are dissolved in about 100-150 ml of dry dichloromethane This solution is cooled to 0-5 °C and 1,1 equivalent of *m*-CPBA (85 %) is added in portions Stirring is continued for 4 hours at room temperature after which the *m*-CBA is filtered off The filtrate is washed several times with dichloromethane The remaining solution is then evaporated *in vacuo* (T<30 °C)

General procedure for the preparation of N-hydroxyaminoacid amides 8

The solid oxaziridines **7** were not purified due to their instability but directly dissolved in about 150 ml of methanol and 1-1 equivalents of H₂NOH HCl are added After stirring at room temperature for about 5-12 hours, the solution is triturated with about 1 L of dry ether The precipitate is isolated by filtration

General procedure for the preparation of the amides 10

The hydrochlorides of **9**(20 mmol) are suspended in 150 ml CHCl₃, together with 1 equivalent(2.03 g, 20 mmol) of triethylamine After 15 minutes the solvent is removed, the residue extracted with diethylether(3x) The combined etherfractions are concentrated *in vacuo* yielding the free α -amino ester The yields are almost quantitative, except in the case of the hydrochloride of alanine ethyl ester **9a**(16%) which has been handled in another way(*vide infra*) The free α -amino ester is immediately dissolved(to prevent self aminolysis) in 140 ml 40% CH₃NH₂/H₂O After half an hour the reaction mixture is concentrated *in vacuo*, yielding amides **10** in almost quantitative yields(see table 2) In the case of **9a** the hydrochloride was dissolved immediately in 40% CH₃NH₂/H₂O After half an hour the reaction mixture was concentrated *in vacuo* yielding a residue containing equimolar amounts of **10a** and CH₃NH₂ HCl This residue is used without further purification for the preparation of **11a**

General procedure for the preparation of the isoxazolidinones 11

Twenty mmol amide **10** and 20 mmol freshly distilled *p*-methoxybenzaldehyde are dissolved in 150 ml of MeOH This solution is refluxed over molecular sieves 3 Å during 18 hours after which the solvent is evaporated The residue is purified by column chromatography

11a The residue containing equimolar quantities of amide **10a** and CH₃NH₂ HCl(see general procedure for amides **10**) are refluxed in methanol with *p*-methoxybenzaldehyde as described above The residue is purified chromatographically(eluent 2% MeOH/CH₂Cl₂) The product **11a** is a mixture of two diastereomers(R_f 0.50, 3% MeOH/CH₂Cl₂)

11b: According to TLC, the diastereomers are formed in a ratio of about 1/1(1% MeOH/CHCl₃) The residue is subjected to flash column chromatography(eluent 1% MeOH/CH₂Cl₂) The eluate is divided in three fractions, the first yielding pure diastereomer II(R_f 0.52, 3% MeOH/CHCl₃)(crystals from CH₂Cl₂/hexane), the last one yielding pure diastereomer I(R_f 0.45, 3% MeOH/CHCl₃) The fraction in between was a mixture of both diastereomers

11c Chromatography of the residue(eluent EtOAc/hexane 40/60) gives two fractions, one

containing diastereomer I (R_f 0.34, EtOAc/hexane 1/1) (crystals from Et₂O/hexane), another containing diastereomer II (R_f 0.11, EtOAc/hexane 1/1) (crystals from CH₂Cl₂/hexane)

11d Chromatography of the residue (eluent 1% MeOH/CH₂Cl₂) gives two fractions, one containing diastereomer I (R_f 0.21, 1% MeOH/CHCl₃) (crystals from CH₂Cl₂/hexane), another containing diastereomer II (R_f 0.11, 1% MeOH/CHCl₃) (crystals from Et₂O/hexane)

General procedure for the preparation of the 1-hydroxy-isoxazolidinones 12

A solution of 1.014 g (5 mmol) of 85% *m*-CPBA in 10 ml CH₂Cl₂ is added dropwise to a cooled solution (ice/water) of **11** (5 mmol) in 100 ml CH₂Cl₂. After 3 hours (unless otherwise stated, see below) the solvent is evaporated and the residue is chromatographed.

12a A mixture of diastereomers of **11a** (see preparation of **11a**) is oxidized with *m*-CPBA. The reaction period has to be extended to 2 days (0°C). TLC showed two iodine positive spots (R_f 0.57 and 0.51, toluene/ethylformate/formic acid 10/7/3). After chromatography of the residue (eluent toluene/ethylformate/formic acid 90/7/3), only one of the diastereomers of **12a** is isolated (24%, R_f 0.51 toluene/ethylformate/formic acid 10/7/3) (crystals from CH₂Cl₂/hexane).

12b A mixture of diastereomers of **11b** (see preparation of **11b**) is oxidized with *m*-CPBA. The reaction period had to be extended to 2 days (0°C). After chromatography (eluent toluene/ethylformate/formic acid 90/7/3) two fractions were collected each containing one of the diastereomers of **12b**. Diastereomer I (14%) R_f 0.49 EtOAc/hexane 1/1 (crystals from CH₂Cl₂/hexane). Diastereomer II (2.5%) R_f 0.32 EtOAc/hexane 1/1 (crystals from CH₂Cl₂/hexane). Another impure fraction containing diastereomer I together with the concentrated filtrates of the previous crystallizations were chromatographed again (eluent EtOAc/hexane 20/80 changing to 50/50). Again two fractions were collected each containing one of the diastereomers. After crystallizations (from CH₂Cl₂/hexane) 13% of diastereomer I and 6.5% of diastereomer II could be isolated. Another impure fraction was chromatographed again (eluent EtOAc/hexane 1/1) and afforded in 7% yield the cyclic nitron **15b** (R_f 0.22, EtOAc/hexane 1/1).

12c Diastereomer II of **11c** was oxidized. The residue was chromatographed (eluent toluene/ethylformate/formic acid 90/7/3), 88% of diastereomer II of **12c** could be isolated (R_f 0.63 toluene/ethylformate/formic acid 10/7/3) (crystals from CH₂Cl₂/hexane).

12d Diastereomer II of **11d** was oxidized. The residue was chromatographed (eluent toluene/ethylformate/formic acid 90/7/3). 74% of diastereomer II of **12d** could be isolated (R_f 0.53 3% MeOH/CHCl₃) (crystals from CH₂Cl₂/hexane).

General procedure for the preparation of the N-hydroxyamides 13

2 mmol of compound **12** is dissolved in 7 N HCl/EtOH together with 139 mg (2 mmol) of H₂NOH·HCl. After 3 hours of *gently* refluxing, the solvent is evaporated. The residue is stripped three times with EtOH. Subsequently the residue is subjected to flash column chromatography.

13a Diastereomer I of **12a** is used for the hydrolysis using hydroxylamine hydrochloride. Eluent 3% MeOH/CH₂Cl₂ changing to 7% MeOH/CH₂Cl₂. Crystals from MeOH/CH₂Cl₂/hexane in the ratio 1/25/25. To check the optical purity of this compound, the compound was synthesized also from optically pure N-hydroxy-alanine ethyl ester^{6a} (3 mmol) and 40% H₂NCH₂/H₂O (1.5 ml) at 0°C. After 1 hour the reaction mixture was concentrated *in vacuo* and stripped with MeOH three times. The residue was crystallized from MeOH/CH₂Cl₂/hexane 1/25/25. Yield 25%. The latter method yielded **13a** with $[\alpha]_D^{20} +45.8$ (c 2, MeOH). **13a** obtained by the oxidative method showed $[\alpha]_D^{20} -45.7$ (c 2, MeOH), so the optical purity is 100%.

13b Diastereomer I of **12b** is used for the hydroxylaminolysis. Eluent 2% MeOH/CH₂Cl₂ changing to 5% MeOH/CH₂Cl₂. Crystals from EtOAc/hexane.

13c Diastereomer II of **12c** is used for the hydroxylaminolysis. Eluent 2% MeOH/CH₂Cl₂ changing to 5% MeOH/CH₂Cl₂. Crystals from CHCl₃/hexane.

13d Diastereomer II of **12d** is used for the hydroxylaminolysis. Eluent 5% MeOH/CH₂Cl₂. Crystals from CHCl₃/hexane.

Spectroscopic data and elemental analyses

6a: (*R*)-isomer

m.p. 115-115.5 °C ¹H-NMR (CDCl₃) δ 0.92 (dd, 6H, CHCH(CH₃)₂), 2.28 (m, 1H, CHCH(CH₃)₂), 3.54 (d, 1H, CHCH(CH₃)₂), 3.81 (s, 3H, PhOCH₃), 5.86 and 6.67 (2H, CONH₂), 6.91 and 7.68 (d 2x, 4H, C₆H₄), 8.05 (s, 1H, N=CHPhOCH₃). IR (KBr, cm⁻¹) 3320 (m), 3180 (m), 1650 (s), 1605 (s). ¹³C-NMR (CDCl₃) δ 17.5 and 19.5 (CHCH(CH₃)₂), 32.8 (CHCH(CH₃)₂), 55.4 (OCH₃), 79.2 (CHCH(CH₃)₂), 114.0, 128.8 and 162.0 (C₆H₄), 161.5 (N=CHPhOCH₃), 175.7 (CONH₂). $[\alpha]_{20}^{20}$

-8 7(c 2, MeOH)

6b: (S)-enantiomer

m p 126-128 °C $^1\text{H-NMR}(\text{CDCl}_3)$ δ 0.90(d, 6H, $\text{CHCH}_2\text{CH}(\text{CH}_3)_2$), 1.52(m, 1H, $\text{CHCH}_2\text{CH}(\text{CH}_3)_2$), 1.76(t, 2H, $\text{CHCH}_2\text{CH}(\text{CH}_3)_2$), 3.92(4H, $\text{CHCH}_2\text{CH}(\text{CH}_3)_2$ and PhOCH_3), 5.43 and 6.66(2H, CONH_2), 6.93 and 7.68(d 2x, 4H, C_6H_4), 8.12(s, 1H, $\text{N}=\text{CHPhOCH}_3$) IR(KBr, cm^{-1}) 3340(m), 3150(m), 1650(s), 1608(s) $^{13}\text{C-NMR}(\text{CDCl}_3)$ δ 21.2, 23.4 and 24.2($\text{CHCH}_2\text{CH}(\text{CH}_3)_2$), 43.6($\text{CHCH}_2\text{CH}(\text{CH}_3)_2$), 55.4(PhOCH_3), 72.2($\text{CHCH}_2\text{CH}(\text{CH}_3)_2$), 114.3, 128.5, 130.0 and 162.2(C_6H_4), 116.3($\text{N}=\text{CHPhOCH}_3$), 176.5(CONH_2) $[\alpha]_{\text{D}}^{19.5}$ -32.5(c 2, MeOH)

6c: (R)-enantiomer

m p 124-5°C $^1\text{H-NMR}(\text{CDCl}_3)$ δ 3.82(s, 3H, OCH_3), 4.90(s, 1H, CHPh), 6.02 and 7.04(2H, NH_2), 6.90-7.75(9H, CHPh , $\text{N}=\text{CHPhOCH}_3$), 8.21(s, 1H, NCHPhOCH_3) IR(KBr, cm^{-1}) 3300(m), 1665(s), 1605(s) $^{13}\text{C-NMR}(\text{CDCl}_3)$ δ 55.4($\text{N}=\text{CHPhOCH}_3$), 76.8(CHPh), 114.1, 128.6, 130.0 and 162.2($\text{N}=\text{CHPhOCH}_3$), 127.2, 127.8, 128.4 and 139.5(CHPh), 162.4($\text{N}=\text{CHPhOCH}_3$), 174.4(CONH_2) $[\alpha]_{\text{D}}^{20}$ +30.0(c 2, MeOH)

6d: (R)-enantiomer

m p 130-2 °C $^1\text{H-NMR}(\text{CDCl}_3)$ δ 2.9-3.38(m, 2H, CH_2Ph), 3.80(s, 3H, $\text{N}=\text{CHPhOMe}$), 4.87(m, 1H, CHCONH_2), 5.84(1H, CHCONH_2 (1H)), 6.90-7.6(11H, CH_2Ph , $\text{N}=\text{CHPhOMe}$ and NH_2 (1H)) IR(KBr, cm^{-1}) 3350(m), 3160(m), 1660(s), 1605(s) $^{13}\text{C-NMR}(\text{CDCl}_3)$ δ 41.1(CHCH_2Ph), 55.3($\text{N}=\text{CHPhOMe}$), 74.9(CHCONH_2), 114.0, 128.2 and 130.0($\text{N}=\text{CHPhOMe}$), 126.4, 128.1, 129.8 and 137.6(CHCH_2Ph), 162.0(CHCONH_2), 175.0($\text{N}=\text{CHPhOMe}$) $[\alpha]_{\text{D}}^{21}$ -262.5(c 2, MeOH)

6e: racemic modification

m p 136-8°C $^1\text{H-NMR}(\text{CDCl}_3)$ δ 1.24 and 1.43(m 2x, 2H, $\text{CHCH}_2\text{CH}_2\text{Ph}$), 1.77(m, 2H, $\text{CHCH}_2\text{CH}_2\text{Ph}$), 2.92(m, 1H, $\text{CHCH}_2\text{CH}_2\text{Ph}$), 3.00(s, 3H, $\text{N}=\text{CHPhOMe}$), 4.66 and 5.86(2H, CHCONH_2), 6.1-6.9(m, 9H, $\text{N}=\text{CHPhOMe}$ and $\text{CHCH}_2\text{CH}_2\text{Ph}$), 7.23(s, 1H, NCHPhOMe) IR(KBr, cm^{-1}) 3310(m), 3150(m), 1650(s), 1605(s) $^{13}\text{C-NMR}(\text{CDCl}_3)$ δ 31.8 and 36.2($\text{CHCH}_2\text{CH}_2\text{Ph}$), 55.4($\text{N}=\text{CHPhOMe}$), 114.2, 130.0 and 162.3($\text{N}=\text{CHPhOMe}$), 125.9, 128.4 and 141.1($\text{CHCH}_2\text{CH}_2\text{Ph}$), 162.1($\text{N}=\text{CHPhOMe}$), 175.8(CHCONH_2)

8a: (R)-enantiomer hydrogenchloride

Yield 76 % m p 108-9 °C $^1\text{H-NMR}(\text{DMSO})$ δ 0.94(d, 6H, $\text{CH}(\text{CH}_3)_2$), 1.74(d, 1H, $\text{CH}(\text{CH}_3)_2$), NH_2 not detected IR(KBr, cm^{-1}) 3360(m), 3060(m), 1690(s), 1605(m), 1400(s) $[\alpha]_{\text{D}}^{20}$ -73(c 1, H_2O)

8b: (S)-enantiomer, hydrogenchloride

Yield 65 % m p 114-5°C $^1\text{H NMR}(\text{CDCl}_3)$ δ 0.94(d, 6H, $\text{CH}_2\text{CH}(\text{CH}_3)_2$), 1.29-1.84(m, 3H, $\text{CH}_2\text{CH}(\text{CH}_3)_2$), 3.50(4 lines, X-part of ABX, $\text{CHCH}_2\text{CH}(\text{CH}_3)_2$), 3.8-5.1(s, broad, 2H, NH_2), 7.00-7.54(2H, CONH_2) IR(KBr, cm^{-1}) 3350(m), 3150(m), 1680(s), 1600(s)

8c: racemic modification

Yield 83 % m p 162-3 °C $^1\text{H-NMR}(\text{DMSO})$ δ 4.32(d, 1H, CHPh , $J(\text{H}_\alpha, \text{NH})=8$ Hz), 6.10(d, CHNH_2), 7.45(d, 1H, NH_2 , $J(\text{NH}_2, \text{OH})=2$ Hz), 7.14-7.42(2H, CONH_2), 7.24-7.4(5H, Ph) IR(KBr, cm^{-1}) 3350(m), 3140(m), 1650(s), 1600(m)

8c: (R)-enantiomer hydrogenchloride $[\alpha]_{\text{D}}^{20}$ -107.5(c 1, MeOH)

8c: (S)-enantiomer hydrogenchloride $[\alpha]_{\text{D}}^{20}$ +107.5(c 1, MeOH)

8c: (R)-enantiomer $[\alpha]_{\text{D}}^{20}$ -57.6(c 1, MeOH)

8c: (S)-enantiomer $[\alpha]_{\text{D}}^{20}$ +57.3(c 1, MeOH) m p 137.4-137.8 °C

8d: racemic modification

Yield 80 % m p 145-6°C $^1\text{H-NMR}(\text{DMSO})$ δ 2.62 - 2.77(2H, CHCH_2Ph , $J_{\text{ax}}=8$ Hz, $J_{\text{ab}}=14$ Hz, $J_{\text{bx}}=5$ Hz), 3.45(m, 1H, CHCH_2Ph , $J_{\text{ax}}=8$ Hz, $J_{\text{bx}}=5$ Hz), 5.59(1H, NH , $J_{\text{NH}_2, \text{OH}}=3$ Hz), 7.41(1H, OH , $J=3$ Hz), 7.01 and 7.15(2H, CONH_2), 7.1-7.3(5H, Ph) IR(KBr, cm^{-1}) 3345(m), 3170(m), 1645(s), 1610(s)

8d: (*R*)-enantiomer $[\alpha]_D^{20} +4.3$ (c 1, MeOH)

8d: (*S*)-enantiomer $[\alpha]_D^{20} -4.3$ (c 1, MeOH)

8e: racemic modification, hydrogenchloride m p 128-30 °C IR(KBr, cm^{-1}) 3300(m), 3150(m), 1675(s), 1620(m), 1600(?)

11a (two diastereomers)

$^1\text{H-NMR}(\text{CDCl}_3)$ δ 1.35(d, 3H, CHCH_3), 1.44(d, 3H, CHCH_3), 1.77(s, 1H, NH), 2.62(s, 3H, NCH_3), 2.67(s, 3H, NCH_3), 3.60(q, 1H, CHCH_3), 3.83(s, 3H, PhOMe), 3.84(q, 1H, CHCH_3), 5.18(s, 1H, CHPhOMe), 5.28(s, 1H, CHPhOMe), 6.69 and 7.02(AB, 4H, CHPhOMe , $J_{ab}=8.5$ Hz), 6.73 and 7.02(AB, 4H, CHPhOMe , $J_{ab}=8.6$ Hz) IR(neat, cm^{-1}) 3450(w), 3330(m), 1690(s), 1615(s), 1590(m) MS(EI, m/z) 220(M^+ , 34), 219(66), 163(53), 162(53), 148(100), 113(55)

11b (diast I)

$^1\text{H-NMR}(\text{CDCl}_3)$ δ 0.99(d, 6H, $\text{CH}_2\text{CH}(\text{CH}_3)_2$), 1.23-2.14(m, 4H, $\text{CH}(\text{NH})\text{CH}_2\text{CH}(\text{CH}_3)_2$), 2.64(s, 3H, NCH_3), 3.55(4 lines, X-part of ABX, 1H, NH_2CHCH_2 , $J_{ax}+J_{bx}=12.9$ Hz), 3.86(s, 3H, CHPhOMe), 5.18(d, 1H, CHPhOMe , $J=1.1$ Hz due to coupling with the C_α -proton), 6.97 and 7.29(AB, 4H, CHPhOMe , $J_{ab}=8.7$ Hz) MS(EI, m/z) 262(M^+ , 11), 261(36), 206(63), 205(73), 177(14), 162(100), 149(73), 148(93), 135(30)

11b (diast II)

m p 118.5-119.5 °C $^1\text{H-NMR}(\text{CDCl}_3)$ δ 0.91 and 0.98(2x d, 6H, $\text{CHCH}_2\text{CH}(\text{CH}_3)_2$), 1.22-2.03(m, 3H, $\text{CHCH}_2\text{CH}(\text{CH}_3)_2$), 1.85(s, 1H, NH), 2.66(s, 3H, NCH_3), 3.73(m, 1H, $\text{CHCH}_2\text{CH}(\text{CH}_3)_2$), 3.83(s, 3H, CHPhOMe), 5.24(d, 1H, CHPhOMe , $J=1.2$ Hz due to coupling with C_α -proton), 6.97 and 7.29(AB, 4H, CHPhOMe , $J_{ab}=8.7$ Hz) IR(KBr, cm^{-1}) 3310(m), 1685(s), 1615(m), 1590(w) MS(EI, m/z) 262(M^+ , 18), 261(39), 206(60), 205(100), 177(10), 162(49), 149(51), 148(56), 135(11) Elem anal calc C 68.67, H 8.45, N 10.68, found C 68.65, H 8.47, N 10.56

11c (diast I)

m p 87.5-88.5 °C $^1\text{H-NMR}(\text{CDCl}_3)$ δ 1.80(s, 1H, NH), 2.67(s, 3H, NCH_3), 3.84(s, 3H, CHPhOMe), 4.88(d, 1H, NHCHPh), 5.44(d, 1H, CHPhOMe , $J=1.5$ Hz, due to coupling with C_α -proton), 6.96 and 7.30(AB, 4H, CHPhOMe , $J_{ab}=8.7$ Hz), 7.26-7.62(m, 5H, CHPh) IR(KBr, cm^{-1}) 3365(w), 3335(w), 1685(s), 1610(m), 1590(w) MS(EI, m/z) 282(M^+ , 21), 225(100), 210(10), 175(7), 148(37), 106(15) Elem anal calc C 72.32, H 6.43, N 9.92, found C 72.35, H 6.43, N 9.85

11c (diast II)

m p 119.5-120.5 °C $^1\text{H-NMR}(\text{CDCl}_3)$ δ 1.92(s broad, 1H, NH), 2.68(s, 3H, NCH_3), 3.84(s, 3H, CHPhOMe), 4.69(d, 1H, NHCHPh), 5.35(d, 1H, CHPhOMe , $J=1.5$ Hz, due to coupling with the C_α -proton), 6.97 and 7.37(AB, 4H, CHPhOMe , $J=8.8$ Hz) IR(KBr, cm^{-1}) 3360(s), 1690(s), 1610(s), 1590(m) MS(EI, m/z) 282(M^+ , 32), 225(100), 210(16), 196(7), 175(7), 148(54), 106(17) Elem anal calc C 72.32, H 6.43, N 9.92, found C 72.40, H 6.44, N 9.84

11d (diast I)

$^1\text{H-NMR}(\text{CDCl}_3)$ δ 1.85(s, 1H, NH), 2.55(s, 3H, NCH_3), 2.93 and 3.10(8 lines, AB-part of ABX, 2H, CHCH_2Ph , $J_{ax}=7.7$ Hz, $J_{bx}=3.7$ Hz, $J_{ab}=13.6$ Hz), 3.78(s, 3H, CHPhOMe), 4.05(X-part of ABX, 1H, CHCH_2Ph , $J_{ax}+J_{bx}=11.1$ Hz), 4.80(d, 1H, CHPhOMe , $J=1.4$ Hz, due to coupling with the C_α -proton), 6.86 and 7.12(AB, 4H, CHPhOMe , $J_{ab}=8.5$ Hz), 7.26(s, 5H, CHCH_2Ph) IR(KBr, cm^{-1}) 3320(w), 1690(s), 1610(m) MS(EI, m/z) 296(M^+ , 6), 205(100), 150(17), 91(10) Elem anal calc C 72.95, H 6.80, N 9.45, found C 72.81, H 6.68, N 9.37

11d (diast II)

$^1\text{H-NMR}(\text{CDCl}_3)$ δ 1.82(s, 1H, NH), 2.53(s, 3H, NCH_3), 3.12 and 3.23(8 lines, AB-part of ABX, 2H, $J_{ax}=4.8$ Hz, $J_{bx}=5.1$ Hz, $J_{ab}=14.0$ Hz, CHCH_2Ph), 3.78(s, 3H, CHPhOMe), 3.84(X-part of ABX, $J_{ax}+J_{bx}=9.6$ Hz, 1H, CHCH_2Ph), 5.06(d, 1H, CHPhOMe , $J=1.0$ Hz, due to coupling with the C_α -proton), 6.73(s, 4H, CHPhOMe), 7.24(s, 5H, CHCH_2Ph) IR(KBr, cm^{-1}) 3320(m), 1680(s), 1610(m) MS(EI, m/z) 296(M^+ , 6), 205(100), 150(16), 91(10) Elem anal calc C 72.95, H 6.80, N 9.45, found C 72.86, H 6.82, N 9.43

12a (one diastereomer isolated)

m p 179.5-180.5 °C $^1\text{H-NMR}(\text{CDCl}_3)$ δ 1.42(d, 3H, CHCH_3), 2.54(s, 3H, NCH_3), 3.53(q, 1H, CHCH_2), 3.82(s, 3H, CHPhOMe), 4.87(s, 1H, CHPhOMe), 5.07(s, broad, 1H, NOH), 6.94 and 7.32(AB, 4H, CHPhOMe , $J_{ab}=8.8$ Hz) IR(KBr, cm^{-1}) 3330(s), 1675(s), 1610(m), 1590(w) MS(EI, m/z) 236(M^+ , 8), 218(8), 162(15), 149(100), 148(100), 128(100) Elem anal calc C 61.00, H 6.83, N 11.86, found C 60.96, H 6.83, N 11.66

12b (diast I)

$^1\text{H-NMR}(\text{CDCl}_3)$ δ 0.96 and 0.99(d 2x, 6H, $\text{CH}(\text{CH}_3)_2$), 1.48-2.17(m, 3H, $\text{CHCH}_2\text{CH}(\text{CH}_3)_2$), 2.55(s, 3H, NCH_3), 3.58(t, 1H, $\text{CHCH}_2\text{CH}(\text{CH}_3)_2$), 3.84(s, 3H, CHPhOMe), 4.88(s, 1H, CHPhOMe), 5.02(s, 1H, NOH), 6.93 and 7.30(AB, 4H, CHPhOMe , $J_{ab}=8.8$ Hz) IR(KBr, cm^{-1}) 3350(s), 1680(s), 1610(m), 1590(w) MS(EI, m/z) 278(M^+ , 5), 221(22), 205(15), 170(50), 149(90), 114(100) Elem anal calc C 64.73, H 7.97, N 10.06, found C 64.75, H 7.97, N 10.00

12b (diast II)

$^1\text{H-NMR}(\text{CDCl}_3)$ δ 0.90 and 0.94(d 2x, 6H, $\text{CHCH}_2\text{CH}(\text{CH}_3)_2$), 1.47-2.05(m, 3H, $\text{CHCH}_2\text{CH}(\text{CH}_3)_2$), 2.78(s, 3H, NCH_3), 3.61(t, 1H, $\text{CHCH}_2\text{CH}(\text{CH}_3)_2$), 3.82(s, 3H, CHPhOMe), 4.8-5.5(s broad, 1H, NOH), 5.36(s, 1H, CHPhOMe), 6.92 and 7.20(AB, 4H, CHPhOMe , $J_{ab}=8.7$ Hz) IR(KBr, cm^{-1}) 3260(m), 1675(s), 1610(m), 1585(w) MS(EI, m/z) 278(M^+ , 2), 221(20), 205(15), 170(44), 149(84), 114(100) Elem anal calc C 64.73, H 7.97, N 10.06, found C 64.67, H 8.00, N 9.85

12c (diast II)

m p 176-177°C(dec) $^1\text{H-NMR}(\text{CDCl}_3)$ δ 2.62(s, 3H, NCH_3), 3.85(s, 3H, CHPhOMe), 4.54(s, 1H, CHPh), 5.02(s, 1H, NOH), 5.06(s, 1H, CHPhOMe), 6.98 and 7.54(AB, 4H, CHPhOMe , $J_{ab}=8.7$ Hz), 7.31-7.54(m, 5H, CHPh) IR(KBr, cm^{-1}) 3410(s, broad), 1695(s), 1610(m), 1585(w) MS(EI, m/z) 298(M^+ , 6), 280(31), 224(18), 190(100), 149(84), 148(82) Elem anal calc C 68.44, H 6.08, N 9.39, found C 68.32, H 6.04, N 9.36

12d (diast II)

m p 160-161 °C $^1\text{H-NMR}(\text{CDCl}_3)$ δ 2.51(s, 3H, NCH_3), 3.04 and 3.28(8 lines, AB-part of ABX, 2H, $J_{ax}=7.0$ Hz, $J_{bx}=4.4$ Hz, $J_{ab}=13.9$ Hz, CHCH_2Ph), 3.80(s, 3H, CHPhOMe), 3.84(X-part of ABX, 1H, CHCH_2Ph), 4.71(s, 1H, NOH), 4.86(s, 1H, CHPhOMe), 6.88 and 7.11(AB, 4H, CHPhOMe , $J_{ab}=8.7$ Hz), 7.26(s, 5H, CHCH_2Ph) IR(KBr, cm^{-1}) 3300(s, broad), 1680(s), 1610(m), 1585(w) MS(EI, m/z) 312(M^+ , 2), 294(6), 221(100), 204(19), 148(30), 91(16), 85(54) Elem anal calc C 69.21, H 6.45, N 8.97, found C 69.17, H 6.49, N 8.81

13a

m p 131-132 °C $^1\text{H-NMR}(\text{CDCl}_3)$ δ 1.25(d, 3H, CHCH_3), 2.85(d, 3H, NHCH_3), 3.60(q, 1H, CHCH_2), 4.8-5.3(s broad, 2H, NHOH), 6.5-6.9(s broad, 1H, NHCH_3) IR(KBr, cm^{-1}) 3100-3500(s, broad), 1650(s), 1550(s) MS(CI, m/z) 119(M^++1 , 77), 103(8), 87(11), 74(17), 60(100) Elem anal calc C 40.67, H 8.53, N 23.71, found C 40.97, H 8.56, N 23.27 $[\alpha]_D^{20}$ -45.7(c 2, MeOH)

13b

m p 85-86°C $^1\text{H-NMR}(\text{CDCl}_3)$ δ 0.94(d, 6H, $\text{CH}_2\text{CH}(\text{CH}_3)_2$), 1.29-1.84(m, 3H, $\text{CH}_2\text{CH}(\text{CH}_3)_2$), 2.86(d, 3H, NHCH_3), 3.50(4 lines, X-part of ABX, $\text{CHCH}_2\text{CH}(\text{CH}_3)_2$, $J_{ax} + J_{bx}=14.4$ Hz), 3.8-5.1(s broad, 2H, NHOH), 6.1-6.6(broad, 1H, NHCH_3) IR(KBr, cm^{-1}) 3340(s), 3250(s), 3130(s), 1660(s), 1560(s) MS(CI, m/z) 161(M^++1 , 100), 141(21), 102(82), 86(14) $[\alpha]_D^{20}$ -28.0(c 2, CHCl_3)

13c

$^1\text{H-NMR}(\text{CDCl}_3)$ δ 2.83(d, 3H, NHCH_3), 4.2-5.0(broad, 2H, NHOH), 4.62(s, 1H, CHPh), 6.4-6.7(broad, 1H, NHCH_3), 7.35(s, 5H, CHPh)

13d

m p 151-152.5 °C $^1\text{H-NMR}(\text{CDCl}_3)$ δ 2.36-3.93(s broad, 2H, NHOH), 2.81 and 3.12(8 lines, AB-part of ABX, 2H, $J_{ax}=9.7$ Hz, $J_{bx}=4.9$ Hz, $J_{ab}=13.9$ Hz, CHCH_2Ph), 2.84(d, 3H, NHCH_3), 3.67(4 lines, X-part of ABX, 1H, $J_{ax} + J_{bx}=14.5$ Hz, CHCH_2Ph), 6.2-6.6(s broad, 1H, NHCH_3), 7.27(s, 5H, CHCH_2Ph) IR(KBr, cm^{-1}) 3390(s), 3220(s), 1645(s), 1550(s) MS(CI, m/z) 195(M^++1 , 52), 120(100), 118(45), 91(51), 58(32) Elem anal calc C 61.84, H 7.27, N 14.42

found C 61.78, H 7.27, N 14.35 $[\alpha]_{\text{D}}^{20} +6.6$ (c 2, MeOH), $[\alpha]_{\text{H}365}^{20} +30.6$ (c 2, MeOH)

15b

$^1\text{H-NMR}$ (CDCl_3) δ 0.96 and 0.98(d 2x, 6H, $\text{CH}_2\text{CH}(\text{CH}_3)_2$), 2.00-2.71(m, 3H, $\text{CH}_2\text{CH}(\text{CH}_3)_2$), 2.91(s, 3H, NCH_3), 3.83(s, 3H, CHPhOMe), 5.70(s, 1H, CHPhOMe), 6.97 and 7.24(AB, 4H, CHPhOMe , $J_{\text{ab}}=8.6$ Hz) MS(EI, m/z) 276(14, M^+), 259(88), 148(100), 135(84), 121(17)

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